

A Histone Cycle: A Systematic Overview on Histone (Octamer Protein) Replication

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ABSTRACT

Genetic information plays an important role in each individual for their differentiation. DNA replication is a highly conserved process that accurately copies the genetic information from one generation to the next. The processes of chromatin disassembly and reassembly during DNA replication also have to be precisely regulated to ensure that the genetic material is compactly packaged to fit into the nucleus while also maintaining the epigenetic information that is carried by the histone proteins bound to the DNA, through cell divisions. So, for the synthesis of genome or gene a group of eight proteins collectively known as octamer proteins or histones plays a crucial role. Colloid of chromosomes is converted into the double stranded DNA structure with intermediate structure of histone proteins. As eukaryotic replication disrupts each nucleosome as the fork passes, followed by incorporation of newly synthesized histones into nucleosomes in the daughter genomes. For the cellular machinery to access the DNA, the chromatin must be unwound and the DNA cleared of histone proteins. In this review, we focus on the process of replication-coupled nucleosome assembly to understand how characteristic steady-state nucleosome landscapes are attained. Recent studies have begun to elucidate mechanisms involved in histone transfer during replication and maturation of the nucleosome landscape after disruption by replication. A fuller understanding of replication-coupled nucleosome assembly will be needed to explain how epigenetic information is replicated at every cell division. We also give details about

regulation of histone in human and the DNA Damage Response.

Keywords: Histone, Chromatin assembly, Histone replication, DNA damage response, Translation and transcription, Histone cycle.

INTRODUCTION

As we know every time when cell divided to form a new cell then there is also DNA (Deoxyribose Nucleic Acid) material get equally distributed into that duplicate cells. [27] In this process DNA replication in cell varies proteins are used involved which are interact with each other. [38] Abnormally in any step of cell division leads to cell death and genetic instability leads to cause disease condition in human. [1] Each step of DNA replication is effectively regulated in the eukaryotic cells. One of the important feature is DNA strongly associated with histone which is a basic protein wrapped around the DNA consist octameric (8 units) basic proteins structure called nucleosomes. [27,28]

The nucleosomes and associated DNA are commonly known as chromatin. [27] Presence of nucleosomes is one of the major principle differences between eukaryotic and prokaryotic DNA replication cause change in whole chromatin structure. [2] Since nucleosomes remove transiently chromatin remodeling depends upon assembly and disassembly of nucleosomes. This chromatin remodeling factors and

protein are able to destabilize interaction between histones and DNA. Hence, the destabilized reaction allows them to interact with other DNA complexes. [3] Nucleosomes are help in DNA packing during S-phase about 30 million nucleosomes are synthesized. [2] Histone production is

coupled with DNA synthesis and shut off process when replication finishes. Histone regulation is important avoid histone accumulation of free histone chromatin is acts as structural barrier and also play important role in regulation. [39]

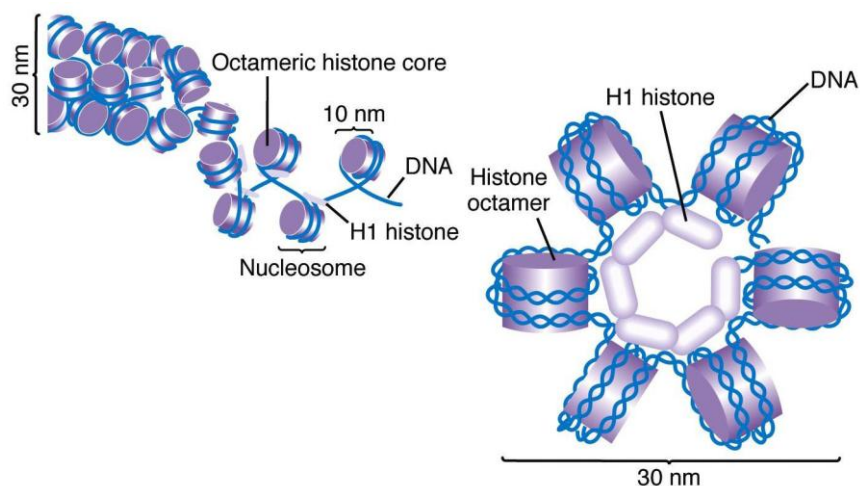


Figure 1 : Histone Octamer^[128]

Histone replication in respect to chromatin: -

Early stage influence of chromatin on replication of histone: -

The replication of DNA requires series of proteins in initial stage in any organism. This series of proteins ultimately leads to loading two hexameric DNA helicases. [40] This protein starts unwinding of DNA which is required for replication. In eukaryotic cells pre-complex formed by two MCM-7 (Minichromosome Maintenance-7 complex) binds to ORC (origin recognition complex) rings. [41] The formed complex is inactive in nature which is activated by Dbf4 (Dumbbell former-4 protein) dependent kinase (DDK) and cyclin dependent kinase (CDK). [4] DNA-replication takes place when DNA sequence contains nucleosome free region (NFR). [5,6,7,8] In drosophila follicle cells histones round's origin of DNA replication initiations (ORIS) are hyper acetylated and change level of acetylation which affects on ORC binding. [9] Methylation of histone H₄ also helpful in

ORC recruitment and promotes ORC binding by methyl transferase SET₈. [10]

When ORC bind to DNA protein CDC₆ (Cell division cycle 6) and CDT (DNA replication factor), this helps to load two MCM2-7 helicases to DNA. [11] This influence by acetylation of H₄ due to recruitment of histone acetyltransferase HBO to ORC by CPT₁ (camptothecin 1) regulation of firing depends on MCM helicase activation by phosphorylation of some subunit by DDK and CDK kinase that allows CDC₄₅ and GINS complex recruitments. [12,13,14] Once this all proteins loaded then replication starts. The spatial arrangement of chromatin helps to define replication domain. [15] The domain containing megabases of contiguous DNA that replicate early than others and this correlated with acetylation levels of H₄. [16,17] The recruitment and activity of replication machinery is influenced by chromatin. When machinery fully set up then replication fork is progressed and DNA starts replicate after nucleosome displacement. [18]

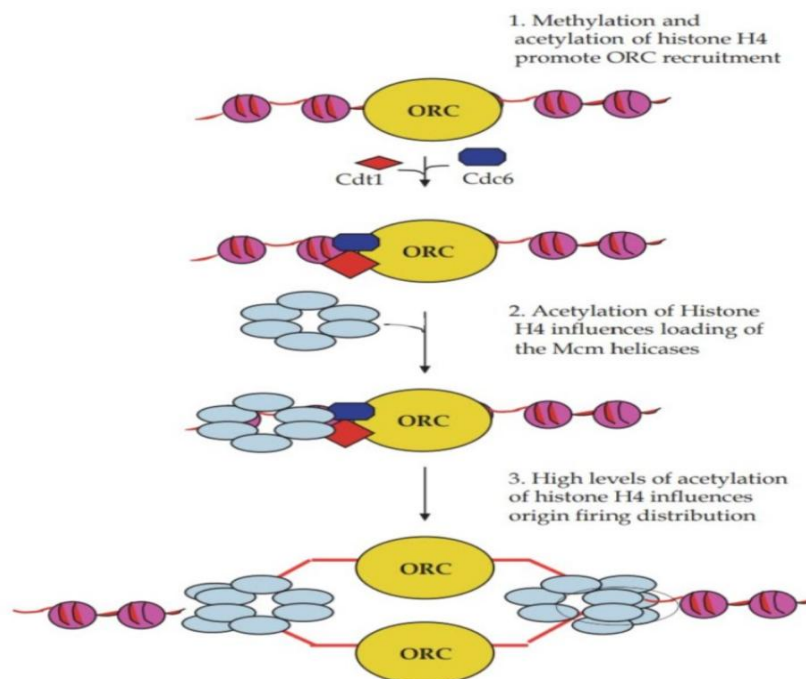


Figure 2 : Early Stage Chromatin Influence on Replication^[32]

Nucleosome arrangement around replication fork: -

During progression of replication fork many proteins and DNA strongly interact with each other. ^[19,20] Chromatin is condensed before replication fork in mammalian cells. ^[21] This condensation process leads to increase mobility of histone H₄ by cyclin A-CDK complete phosphorylation. ^[22] It is still not clear that nucleosome decondensed before replication is due to passage of replication machinery or chromatin remodeling. ^[3] Proteins are able to react with chromatin complete and make them unstable for transcription. After transcription they interact with DNA. ^[42] The major two complexes involved in H₂A and H₂B displacement is FACT (facilitates chromatin transcription complex) and NAP₁K (nucleosome assembly protein kinase) and for H₃/H₄ displacement ASF₁ and CAF₁ present [in this N-terminal of both H₂A and H₂B binds with FACT to form complex with nucleosomes. The mid domain of nucleosomes attached with H₃-H₄ tetramer [(CH₃-CH₄)₂] to displace H₂A-H₂B dimer from nucleosomes. ^[43] During displacement the subunit of FACT complex i.e. Spt₁₆ and SSRP (structure specific

recognition protein), are play important role which shows indirect effect on nucleosome reorganization. ^[23,24] The other NAP (nucleosome assembly protein), complex shows disassembly of nucleosomes when it combined with RSC complex. ^[25,26] ASF₁ and CAF₁ combined to cause deposition of histone H₃/H₄ in passage of replication fork by binding with PCNA (Proliferating cell nuclear antigen), replication factor C and MCM helicase complex due to deposition of histone octamer DNA wrapped around nucleosome. ^[29,30] H₃/H₄ tetramer recycled after each DNA strand replication and new tetramer deposited over other strand of DNA. ^[31] CAF plays role in leading and lagging strand by PCNA. Depletion in CAF₁ cause decrease chromatin replication and increase DNA damage. ^[32,33] Finally, H₁ proteins cause chromatin compaction S-phase progression. ^[34] Due to DNA polymerase polarity on 5`to3` lagging strand chromatin landscape occurs which generate short fragment of DNA known as okazaki fragment. ^[35] This fragment needs to be goes under process of maturation for that flap endonuclease (FEN₁) and DNA ligase-1 plays important roles. This enzyme cause

flap processing and ligation of okazaki fragments. [36,37]

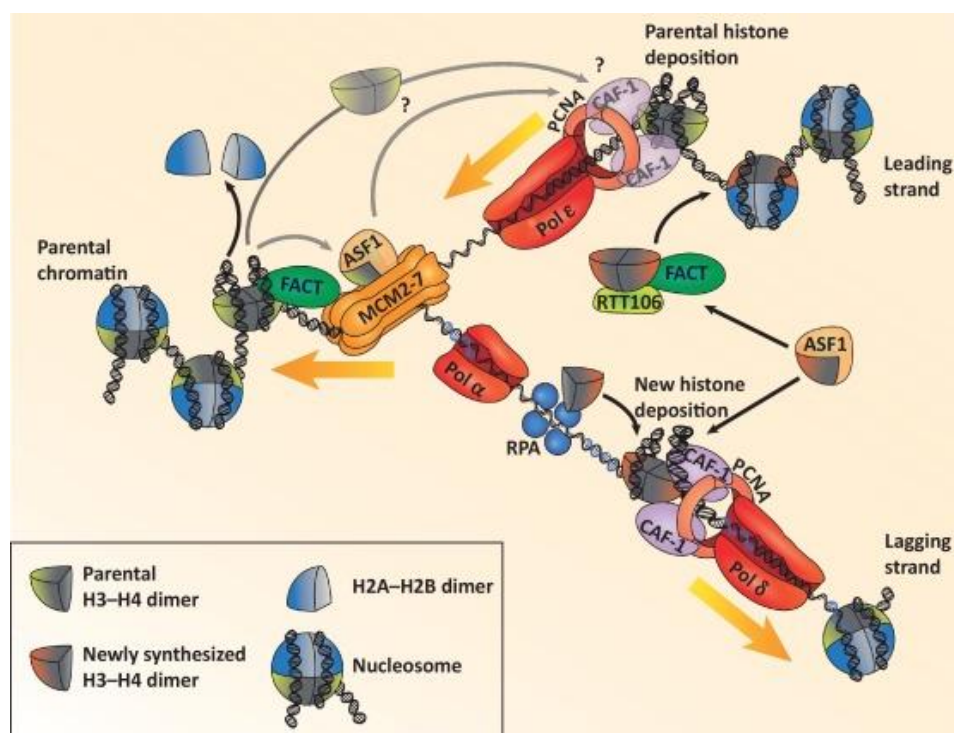


Figure 3 : Nucleosome Rearrangement around Replication Fork [129]

Maturation of chromatin and centromere formation: -

When nucleosome attached to DNA there is two processes are takes place i.e. maturation of chromatin and centromere formation. Histone starts to take some place and causes modification before DNA reposition. [57] Newly formed chromatin is highly acetylated and it need to be deacetylate or methylated to reach compact state. [58] Deacetylation cause due to enzyme deacetylase and methylation cause due to enzyme DNA methyl transferase in compaction state other post-transcriptional modifications are happening and establish some epigenetic code transfer to daughter cells. [54,55] This epigenetic memory is important for differentiation. [56] All processes do not happen in replication some of also takes place in mitosis. Chromatin replication near centromere ensures segregation during mitosis. The heterochromatin named CENP-A (Centromere protein A) helps for binding kinetochore during mitosis. [59] Correct segregation leads to formation of spindle fibers. [53] The highly diversified protein

called heterochromatin (CENP-A) replaces histone H₃ at centromeric DNA. [51,52] This heterochromatin interspersed with canonical nucleosomes and promotes folding of chromatin during metaphase. [49] This formed complex allows kinetochore and microtubule attachment. [49] Heterochromatin positioned on centromeric chromatin during telophase after chromosome separation. [50] Defect in this process cause genetic instability, chromosome loss and cell death. [46,47,48] Several studies show that proper homeostasis between H₃ and heterochromatin (CENP-A) is help for distribution of centromeric variant and chromosomal segregation. [44,45]

Histone regulation: -

Nucleosomes are more than structural bricks for DNA and cause modification on histone to maintain epigenetic state. Hence, DNA replication is interdependent on chromatin reorganization. [61] Highly cell-cycle regulated genes are histone genes because cell generation needs high amount of histone replication. [60] The

histone mainly regulated by transcription, translation and post-transcription way but metabolism depends on organisms. [62,63] The bacterial species like *S. cerevisiae* shows transcriptional regulation. [64] While in the mammalian cells post-transcriptional and translational mechanism seen. [65]

1) Transcriptional regulation of histone: -

It is most important in bacterial species i.e. *S. cerevisiae* regulated in G1 phase of cell cycle or life cycle. [66] In higher eukaryotic it happens all of histone stages. [67] Histone expression must be stoichiometric if imbalance cause then it leads to deterioration of cell or disruption cell. [44,45,68] In S-phase replication expression increased by three to five folds. [69] In each histone cluster consist at least five canonical histones. Which are well organized and co-ordinated octamer-binding transcription factor (OTF₁) for H₂B activates coding region sequence in H₂A, H₃ and H₄ genes. [70,71]

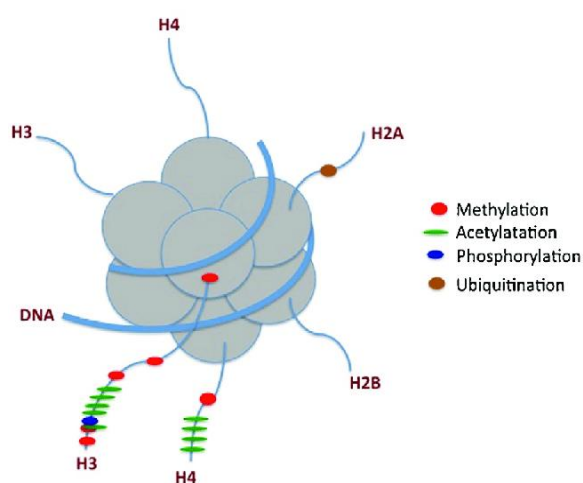


Figure 4 : Histone Modifications in Regulation [130]

Nuclear protein ataxia-telangiectasia's (NPAT) is required for activation of histone gene for S-phase Progression. [72] NPAT locates next to histone locus bodies and undergoes phosphorylation by cyclin E-CDK₂ and increase gene transcription. [73,74] Histone gene express and promotes upstream activating sequences (UAS) for recruitment of Spt₁₀ and SBF. [75] Histone gene also

consist negative regulatory site for replication under stress condition. [76,77,78] The HIR (histone regulator) complex responsible for transcription is formed by proteins named HIRA, ubinuclein₁ and cabin₁. The HIRA associated with chromatin assembly during transcription. [79] Overexpression of HIRA cause blockage of transcription in S-phase HIRA phosphorylated by cyclin E-CDK₂ and cyclin A-CDK₂. [80,81]

2) Post-transcriptional and translational regulation of histones: -

Mammalian histone mRNA contains 3' UTR (untranslated region) sequence for loop formation but they lack of introns and poly (A) tail. [69] The transcribed histone localize to cajal bodies which express NPAT and mature U₇snRNA of histone locus bodies. [82] Maturation occurs after 3' end formation by endonucleolytic cleavage. This cleavage occurs between stem loop and histone downstream element (HDE). [83,84] This cleavage involved by specific proteins like SLBP, LSM₁₋₁₁, U₇snRNA and ZFP₁₀₀. stem-loop binding protein (SLBP) is major protein for translation and post-transcription process for regulation of histones. SLBP is major protein known as cell cycle regulated protein accumulate in G₁ phase and ends in S-phase. [85] SLBP helps to cleavage during mRNA maturation facilitate circulation of histone and translated by polyribosomes and also increase histone mRNA stability and prevent histone degradation when DNA replication inhibits then SLBP also get degraded some of studies shows degradation of H₂B transcript by some component of exosomes. [86,87,88] Length of poly (A) is cell-cycle dependent it decreases in G₁ phase and progress in S-phase and progress in S-phase to G₂ phase. [89]

Regulation of histone by controlled level of protein: -

Studies shows that histone mRNA level controls the histone protein level. [90] The regulation pathway involves action of

CHK₂ and Rad₅₃ which play important role in DNA damage response. [1,91] Degradation of histone involves rad₅₃ action with E₂ ubiquitin ligases. [92] The mechanism of

degradation involves phosphorylation of tyrosine and ubiquitylation before proteolysis histone degradation does not involve kinases. [93]

DNA damage response (DDR): coupling of DNA and synthesis of histone: -

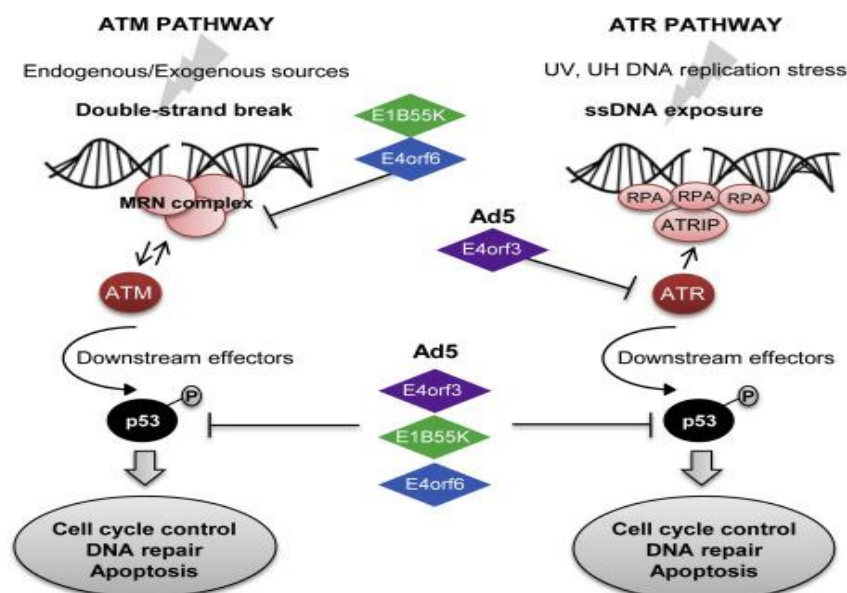


Figure 5 : DNA Damage Response Pathways [131]

DDR is one of the most important checkpoint in cell damage when DNA sense damage. [101] DDR activation leads to action of kinase cascade which leads to correct the cell damage. [102] If DDR is not activated then cell damage will not repair and enters into apoptosis programs and after apoptosis cell will die. [94] Cell cycle involves proper functioning so that it maintains genome integrity and mutations in disease conditions. DDR can resist G₁-phase s-phase and G₂ - Phase. [95] Two enzymes i.e. ATM (ataxia-telangiectasia mutated) and ATR kinase (ATR and Rad3-Related) are important in human cell and Tel₁ and Mec₁ are important enzymes in bacteria S-cerevisiae for activation of DDR. The ATM kinase directly involved in histone expression. [102,103] In human cells NPAT phosphorylation by cyclin E-CDK₂ is required for human gene transcription. In S-phase NPAT activation is important for histone expression. [72] Activation of ATM, P₅₃ and P₂₁ required for synthesis of histone upon DNA damage. [103] E-CDK₂ complex activity resist by enzyme P₂₁. Due to

complex (cyclin E-CDK₂) inhibition the enzyme NPAT is dephosphorylated and activates the transcription. [96] The histone repression cause by change in HIRA activity for or location damage of DNA strands also cause post-transcriptional degradation of histone mRNAs. [104] Histone m RNA undergoes oligouridylation while cell treat with hydroxyl urea (HU). [97] This oligouridylation of m RNA histone depends on UPF₁ which binds to SLBP to recruit 3' terminal uridylyl transferase (TUT-ase). The 3' oligo (U) tails triggers the Lsm₁₋₇ complex for progression of m RNA degradation by exosomes and Xrn₁. [97] Rad₅₃ pathway not directly plays as part of DDR but it could be important for important for destruction of translated histones. Studies shows that Asf₁ and Rad₅₃ plays important role in histone repression process. [105] The stable complex of Asf₁ and Rad₅₃ is dissociate by phosphorylation due to Mec₁ phosphorylate in activation of DDR. [106] DNA repair involves DDR checkpoint activation and chromatin remodeling in which Asf₁ plays crucial role. [98] The other

important enzyme named Rad₅₃ can available in G₁, G₂ and M-phase in hypophosphorylated form which is depend on Cdc₂₈ (A homologue of human CDK₁ and CDK₂). HIR complex will leads to form precipitation with Asf₁. This leads to nucleosomes assembly and DNA template. [99] Asf₁ lacking in mRNA cause abnormalities in S-phase. [100] The dissociation of Asf₁ – Rad₅₃ complex is important in histone transcription. [105]

Free histone formation: -

During a normal cell cycle DNA replication is unbalance due to histone supply unbalancing. [114] There are two possible scenarios for histone supply in which first occurs due to difference between rate of DNA synthesis and supply of histone during DNA replication. [107,108] More replication forks are used in S-phase and also replication stress affect on speed of replication fork. [109,110,111,112] In second situation cell can use free histones during G₂-phase of cell cycle. All free histones should be degrading after balanced ratio of histone H₃ and CENPA cause finished replication by chromosomes segregation. [104,116] So, here we can say that abnormality or imbalance of above two histone types (H₃ and CENPA) taken active participation in replication of cells which will lead to high incidence rate of chromosomes loss. [113]

Free histone transcription: -

Assemble and disassembly of nucleosomes is also required in transcription of chromatin template after passing of RNA polymerase II (RNA Pol-II). This transcribed chromatins are the main source of free histone formation. Due to imbalance between histone supply and demand the free histone amount is rises. The FACT complex is mainly involved in the transcription process along with RNA polymerase-II. [116,117] FACT complex stimulates the enzyme RNA Pol-II for elongation process in transcription. [118,119,120,121,122] FACT complex linked with H₃/H₄ tetramer and H₂A and H₂B dimer shows integrity towards

one of the subunit. [123,124] Spt₁₆ is play important role in reassembly of H₃ and H₄ histones. [125] Chromatin dysfunction cause generation of Spt₁₆ proteins leads to free histone accumulation. This combined with Rad₅₃K²²⁷A cause degradation of histone and increase the amount of free histones. [126] H₂A-H₂B expression causes suppression of this mutants (Rad₅₃K²²⁷A) and promotes expression of H₂A-H₂B mutation of Spt₁₆ and histone level having correction. [126] The another protein factor called Spt₆ is helping in H₃-H₄ reposition during transcription and having strong negative interaction with mutant i.e. Rad₅₃K²²⁷A. free histones forms due to chromatin reassembly detects during transcription with involvement of Rad₅₃ which negatively interact with proteins involved in chromatin related processes. [127] Some of this factors are involved during chromatin related transcription processes. [127,126]

CONCLUSION

As we know the DNA is helpful for transmission of genetic information from one generation to another generation in the form of genomics or genetic code which having a particular sequence of genome in it. This topic highlights the histone cycle which is main protein in DNA transcription process. As we know the DNA is mainly found in eukaryotic organisms but it also found in some of the aracheabacterial species. (B. Thermophiles) which is the oldest colonies of bacteria living on earth. In this DNA forms a nucleoprotein complex called chromatin helps in compaction of genomic DNA in smaller space of nucleus. Hence, the chromatin works as a building block in nucleosome formation. The cell has its own unique and complex machinery to carried out various processes involving DNA needs to modify chromatin first. Chromatin mainly acts as a regulation machinery which carries epigenetic code which are important as one of contained of DNA in cell. In higher organisms with large and complex genome the particular cell require particular fraction of genome is

active. For this a brief passage of replication fork around replication coupled assembly is required. Histone synthesis is coupled to replication of genomic DNA and the existent of specific factor CAF-1 (chromatin activating factor-1) which targets newly synthesized histones to replicating DNA. The post-translational modifications and protein degradation may be involved in regulating the activity CAF-1 in a cell-cycle-specific manner. Outside replication fork nucleosome assembly independent on histone H₃ variant and replication-independent pathways. Apart from an inherent of replication –coupled assembly to impose basal silencing, inheritance of post translationally modified histones at silent domains could play additional roles in maintenance of silent states. This topic also focuses on balance level of histones during chromatin formation and avoids deleterious effects due to generation of free histones. As a potential carrier of epigenetic information the recycling of parental histones after the passage of the replication fork is important for the inheritance of chromatin states. Recent structure studies revealed extensive details on the co-operation among replisome (responsible for unwinding of DNA & promotes replication) especially the co-operation among the replicative helicase and DNA polymerase (DNA Pol-II). This evidence strongly supports replisome components that directly contribute into histone cycle.

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REFERENCES

1. Jones RM, Petermann E. Replication fork dynamics and the DNA damage response. *Biochem J.* 2012 Apr 1;443(1):13-26.
2. Gunjan A, Paik J, Verreault A. Regulation of histone synthesis and nucleosome assembly. *Biochimie.* 2005 Jul;87(7):625-635.
3. Alabert C, Groth A. Chromatin replication and epigenome maintenance. *Nat Rev Mol Cell Biol.* 2012 Mar;13(3):153-167.
4. Mechali M. Eukaryotic DNA replication origins: many choices for appropriate answers. *Nat Rev Mol Cell Biol.* 2010 Oct;11(10):728-738.
5. Xu J, Yanagisawa Y, Tsankov AM, et al. Genome-wide identification and characterization of replication origins by deep sequencing. *Genome Biol.* 2012 Apr 24;13(4): R27.
6. Gao F, Luo H, Zhang CT. DeOri: a database of eukaryotic DNA replication origins. *Bioinformatics.* 2012 Jun 1; 28(11):1551-1552.
7. Sclafani RA, Holzen TM. Cell cycle regulation of DNA replication. *Annu Rev Genet.*2007;41:237-280.
8. Dohrmann PR, Sclafani RA. Novel role for checkpoint Rad53 protein kinase in the initiation of chromosomal DNA replication in *Saccharomyces cerevisiae*. *Genetics.* 2006 Sep;174(1):87-99.
9. Aggarwal BD, Calvi BR. Chromatin regulates origin activity in *Drosophila* follicle cells. *Nature.* 2004 Jul 15; 430(6997):372-376.
10. Tardat M, Brustel J, Kirsh O, et al. The histone H4 Lys 20 methyltransferase PR-Set7 regulates replication origins in mammalian cells. *Nat Cell Biol.* 2010 Nov;12(11):1086-1093.
11. Kouzarides T. Chromatin modifications and their function. *Cell.* 2007 Feb 23;128(4):693-705.
12. Miotto B, Struhl K. HBO1 histone acetylase is a coactivator of the replication licensing factor Cdt1. *Genes Dev.* 2008 Oct 1;22(19):2633-2638.
13. Miotto B, Struhl K. HBO1 histone acetylase activity is essential for DNA replication licensing and inhibited by

- Geminin. *Mol Cell*. 2010 Jan 15;37(1):57-66.
14. Iizuka M, Matsui T, Takisawa H, et al. Regulation of replication licensing by acetyltransferase Hbo1. *Mol Cell Biol*. 2006 Feb;26(3):1098-1108.
 15. Remus D, Diffley JF. Eukaryotic DNA replication control: lock and load, then fire. *Curr Opin Cell Biol*. 2009 Dec;21(6):771-777.
 16. Schwaiger M, Stadler MB, Bell O, et al. Chromatin state marks cell-type- and gender-specific replication of the *Drosophila* genome. *Genes Dev*. 2009 Mar 1;23(5):589-601.
 17. Gilbert DM, Takebayashi SI, Ryba T, et al. Space and time in the nucleus: developmental control of replication timing and chromosome architecture. *Cold Spring Harb Symp Quant Biol*. 2010;75:143-153.
 18. Yu-Hung Chen, Sarah Keegan, Malik Kahli, et al. Transcription shapes DNA replication initiation and termination in human cells. *Nature structural & molecular biology*. 2019;26(1):67-77.
 19. Sogo JM, Stahl H, Koller T, et al. Structure of replicating simian virus 40 mini- chromosomes. The replication fork, core histone segregation and terminal structures. *J Mol Biol*. 1986 May 5;189(1):189-204.
 20. Gasser R, Koller T, Sogo JM. The stability of nucleosomes at the replication fork. *J Mol Biol*. 1996 May 3;258(2):224-239.
 21. Kuipers MA, Stasevich TJ, Sasaki T, et al. Highly stable loading of Mcm proteins onto chromatin in living cells requires replication to unload. *J Cell Biol*. 2011 Jan 10;192(1):29-41.
 22. Alexandrow MG, Hamlin JL. Chromatin decondensation in S-phase involves recruitment of Cdk2 by Cdc45 and histone H1 phosphorylation. *J Cell Biol*. 2005 Mar 14;168(6):875-886.
 23. Belotserkovskaya R, Oh S, Bondarenko VA, et al. FACT facilitates transcription-dependent nucleosome alteration. *Science*. 2003 Aug 22;301(5636):1090-1093.
 24. Xin H, Takahata S, Blanksma M, et al. yFACT induces global accessibility of nucleosomal DNA without H2A-H2B displacement. *Mol Cell*. 2009 Aug 14;35(3):365-376.
 25. Ito T, Bulger M, Kobayashi R, et al. *Drosophila* NAP-1 is a core histone chaperone that functions in ATP-facilitated assembly of regularly spaced nucleosomal arrays. *Mol Cell Biol*. 1996 Jun;16(6):3112-3124.
 26. Lorch Y, Maier-Davis B, Kornberg RD. Chromatin remodeling by nucleosome disassembly in vitro. *Proc Natl Acad Sci U S A*. 2006 Feb 28;103(9):3090-3093.
 27. Gerard J. Tortora, Mark T. Nielson. *Principles of Human Anatomy*, 14th edition, willy publication. 2017. p 3,28, 42-:43.
 28. Harsh Mohan. textbook of Pathophysiology, 6th edition, jaypee brother's medical publisher Ltd., 2010. p 23.
 29. Xu M, Long C, Chen X, et al. Partitioning of histone H3-H4 tetramers during DNA replication-dependent chromatin assembly. *Science*. 2010 Apr 2; 328(5974):94-98.
 30. Franco AA, Lam WM, Burgers PM, et al. Histone deposition protein Asf1 maintains DNA replisome integrity and interacts with replication factor C. *Genes Dev*. 2005 Jun 1;19(11):1365-1375.
 31. Groth A, Corpet A, Cook AJ, et al. Regulation of replication fork progression through histone supply and demand. *Science*. 2007 Dec 21;318(5858):1928-1931.
 32. Hoek M, Stillman B. Chromatin assembly factor 1 is essential and couples chromatin assembly to DNA replication in vivo. *Proc Natl Acad Sci U S A*. 2003 Oct 14;100(21):12183-12188.
 33. Ye X, Franco AA, Santos H, et al. Defective S phase chromatin assembly causes DNA damage, activation of the S phase checkpoint, and S phase arrest. *Mol Cell*. 2003 Feb;11(2):341-351.
 34. Finn RM, Browne K, Hodgson KC, et al. sNASP, a histone H1-specific eukaryotic chaperone dimer that facilitates chromatin assembly. *Biophys J*. 2008 Aug;95(3): 1314-1325.

35. Smith DJ, Whitehouse I. Intrinsic coupling of lagging-strand synthesis to chromatin assembly. *Nature*. 2012 Mar 22;483(7390):434-438.
36. Lata Balakrishnan and Robert A. Bambara. Flap Endonuclease 1. *Annu Rev Biochem*. 2013 Jun 2; 82: 119–138.
37. Li Zheng and Binghui Shen. Okazaki fragment maturation: nucleases take centre stage. *J Mol Cell Biol*. 2011 Feb; 3(1): 23–30.
38. Lisa Bartee, Walter Shriener, Catherine Creech. Principles of Biology. OpenStax, Biology. OpenStax CNX. May 27, 2016. p 454,455,460-463.
39. Dorine Rossetto, Nikita Avvakumov, and Jacques Côté. Histone phosphorylation: A chromatin modification involved in diverse nuclear events. *Epigenetics*. 2012 Oct 1; 7(10): 1098–1108.
40. R. A. Sclafani and T. M. Holzen. Cell Cycle Regulation of DNA Replication. *Annu Rev Genet*. 2007; 41:237-280.
41. Susan L. Forsburg. Eukaryotic MCM proteins: beyond replication initiation. *Microbiol Mol Biol Rev*. 2004 mar; 68(1):109-131.
42. Yoav Lubelsky, Joseph A. Prinz, David M. MacAlpine, et al. DNA replication and transcription programs respond to the same chromatin cues. *Genome Res*. 2014 Jul; 24(7): 1102–1114.
43. Tsunaka Y, Tonga J, Yamaguchi H, et al. Phosphorylation intrinsically disordered region of FACT masks its nucleosomal DNA binding elements. *J Biol Chem*. 2009; 284:24610-24621.
44. Castillo AG, Mellone BG, Partridge JF, et al. Plasticity of fission yeast CENP-A chromatin driven by relative levels of histone H3 and H4. *PLoS Genet*. 2007 Jul;3(7): e121.
45. Au WC, Crisp MJ, DeLuca SZ, et al. Altered dosage and mislocalization of histone H3 and Cse4p lead to chromosome loss in *Saccharomyces cerevisiae*. *Genetics*. 2008 May;179(1): 263-275.
46. Choy JS, Mishra PK, Au WC, et al. Insights into assembly and regulation of centromeric chromatin in *Saccharomyces cerevisiae*. *Biochim Biophys Acta*. 2012 Jul;1819(7):776-783.
47. Stoler S, Rogers K, Weitze S, et al. Scm3, an essential *Saccharomyces cerevisiae* centromere protein required for G2/M progression and Cse4 localization. *Proc Natl Acad Sci USA*. 2007 Jun 19;104(25):10571-10576.
48. Dunleavy EM, Roche D, Tagami H, et al. HJURP is a cell-cycle-dependent maintenance and deposition factor of CENP-A at centromeres. *Cell*. 2009 May 1;137(3):485-497.
49. Black BE, Bassett EA. The histone variant CENP-A and centromere specification. *Curr Opin Cell Biol*. 2008 Feb;20(1):91-100.
50. Fujita Y, Hayashi T, Kiyomitsu T, et al. Priming of centromere for CENP-A recruitment by human hMis18alpha, hMis18beta, and M18BP1. *Dev Cell*. 2007 Jan;12(1):17-30.
51. Jansen LE, Black BE, Foltz DR, et al. Propagation of centromeric chromatin requires exit from mitosis. *J Cell Biol*. 2007 Mar 12;176(6):795-805.
52. Schuh M, Lehner CF, Heidmann S. Incorporation of *Drosophila* CID/CENP-A and CENP-C into centromeres during early embryonic anaphase. *Curr Biol*. 2007 Feb 6;17(3):237-243.
53. Westermann S, Drubin DG, Barnes G. Structures and functions of yeast kinetochore complexes. *Annu Rev Biochem*. 2007; 76:563-591.
54. Pesavento JJ, Yang H, Kelleher NL, et al. Certain and progressive methylation of histone H4 at lysine 20 during the cell cycle. *Mol Cell Biol*. 2008 Jan;28(1):468-486.
55. Lanzuolo C, Lo Sardo F, Diamantini A, et al. PcG complexes set the stage for epigenetic inheritance of gene silencing in early S phase before replication. *PLoS Genet*. 2011 Nov;7(11): e1002370.
56. Agustina D'Urso and Jason H. Brickner. Mechanisms of epigenetic memory. *Trends Genet*. 2014 Jun; 30(6): 230–236.
57. Mehta, I.S., Kulashreshtha, M., Chakraborty, S. et al. Chromosome territories reposition during DNA damage-

- repair response. *Genome Biol* 14.2013; R135.
58. Anton Eberharter and Peter B. Becker. Histone acetylation: a switch between repressive and permissive chromatin: Second in review series on chromatin dynamics. *EMBO Rep.* 2002 Mar 15; 3(3): 224–229.
59. Jolien S. Verdaasdonk and Kerry Bloom. Centromeres: unique chromatin structures that drive chromosome segregation. *Nat Rev Mol Cell Biol.* 2011 May; 12(5): 320–332.
60. Marzluff WF, Wagner EJ, Duronio RJ. Metabolism and regulation of canonical histone mRNAs: life without a poly(A) tail. *Nat Rev Genet.* 2008 Nov; 9(11):843–854.
61. Milena Georgieva, Dessislava Staneva and George Miloshev. Epigenetic Significance of Chromatin Organization During Cellular Aging and Organismal Lifespan. *Epigenetics, the Environment, and Children's Health Across Lifespans.* 2016 Feb 9: 21–66.
62. Jing Fan, Kimberly A. Krautkramer, et al. Metabolic Regulation of Histone Post-Translational Modifications. *ACS Chem. Biol.* 2015, 10, 1, 95–108.
63. Diane E. Handy, Rita Castro and Joseph Loscalzo. Epigenetic Modifications: Basic Mechanisms and Role in Cardiovascular Disease. *Circulation.* 2011 May 17; 123(19): 2145–2156.
64. Steven Hahn and Elton T. Young. Transcription regulation in *Saccharomyces cerevisiae*: transcription factor regulation and function, mechanism of initiation and role of activators and coactivators. *Genetics.* 2011 Nov; 189(3): 705–736.
65. N.V. Bhagavan and Chung-Eun Ha. *Essentials of Medical Biochemistry with Clinical Cases.* Second Edition, Elsevier publication, 2015. p 447–464.
66. Amin AD, Vishnoi N, Prochasson P. A global requirement for the HIR complex in the assembly of chromatin. *Biochim Biophys Acta.* 2012 Mar; 1819(3-4):264–276.
67. Jaeger S, Barends S, Giege R, et al. Expression of metazoan replication-dependent histone genes. *Biochimie.* 2005 Sep-Oct; 87(9-10):827–834.
68. Meeks-Wagner D, Hartwell LH. Normal stoichiometry of histone dimer sets is necessary for high fidelity of mitotic chromosome transmission. *Cell.* 1986 Jan 17; 44(1):43–52.
69. Marzluff WF, Duronio RJ. Histone mRNA expression: multiple levels of cell cycle regulation and important developmental consequences. *Curr Opin Cell Biol.* 2002 Dec; 14(6):692–699.
70. Fletcher C, Heintz N, Roeder RG. Purification and characterization of OTF-1, a transcription factor regulating cell cycle expression of a human histone H2b gene. *Cell.* 1987 Dec 4; 51(5):773–781.
71. Kaludov NK, Pabon-Pena L, Hurt MM. Identification of a second conserved element within the coding sequence of a mouse H3 histone gene that interacts with nuclear factors and is necessary for normal expression. *Nucleic Acids Res.* 1996 Feb 1; 24(3):523–531.
72. Ye X, Wei Y, Nalepa G, et al. The cyclin E/Cdk2 substrate p220(NPAT) is required for S-phase entry, histone gene expression, and Cajal body maintenance in human somatic cells. *Mol Cell Biol.* 2003 Dec; 23(23):8586–8600.
73. Ma T, Van Tine BA, Wei Y, et al. Cell cycle-regulated phosphorylation of p220(NPAT) by cyclin E/Cdk2 in Cajal bodies promotes histone gene transcription. *Genes Dev.* 2000 Sep 15; 14(18):2298–2313.
74. Zhao J, Kennedy BK, Lawrence BD, et al. NPAT links cyclin E-Cdk2 to the regulation of replication-dependent histone gene transcription. *Genes Dev.* 2000 Sep 15; 14(18):2283–2297.
75. Eriksson PR, Ganguli D, Clark DJ. Spt10 and Swi4 control the timing of histone H2A/H2B gene activation in budding yeast. *Mol Cell Biol.* 2011 Feb; 31(3):557–572.
76. Spector MS, Raff A, DeSilva H, et al. Hir1p and Hir2p function as transcriptional corepressors to regulate histone gene transcription in the *Saccharomyces cerevisiae* cell cycle. *Mol Cell Biol.* 1997 Feb; 17(2):545–552.

77. Osley MA, Gould J, Kim S, et al. Identification of sequences in a yeast histone promoter involved in periodic transcription. *Cell*. 1986 May 23;45(4):537-544.
78. Freeman KB, Karns LR, Lutz KA, et al. Histone H3 transcription in *Saccharomyces cerevisiae* is controlled by multiple cell cycle activation sites and a constitutive negative regulatory element. *Mol Cell Biol*. 1992 Dec;12(12):5455-5463.
79. Tagami H, Ray-Gallet D, Almouzni G, et al. Histone H3.1 and H3.3 complexes mediate nucleosome assembly pathways dependent or independent of DNA synthesis. *Cell*. 2004 Jan 9;116(1):51-61.
80. Nelson DM, Ye X, Hall C, et al. Coupling of DNA synthesis and histone synthesis in S phase independent of cyclin/cdk2 activity. *Mol Cell Biol*. 2002 Nov;22(21):7459-7472.
81. Hall C, Nelson DM, Ye X, et al. HIRA, the human homologue of yeast Hir1p and Hir2p, is a novel cyclin-cdk2 substrate whose expression blocks S-phase progression. *Mol Cell Biol*. 2001 Mar;21(5):1854-1865.
82. Nizami Z, Deryusheva S, Gall JG. The Cajal body and histone locus body. *Cold Spring Harb Perspect Biol*. 2010 Jul;2(7):a000653.
83. Chodchoy N, Pandey NB, Marzluff WF. An intact histone 3'-processing site is required for transcription termination in a mouse histone H2a gene. *Mol Cell Biol*. 1991 Jan;11(1):497-509.
84. Gu X, Marzluff WF. 3' Processing and termination of mouse histone transcripts synthesized in vitro by RNA polymerase II. *Nucleic Acids Res*. 1996 Oct 1;24(19):3797-3805.
85. Zheng L, Dominski Z, Yang XC, et al. Phosphorylation of stem-loop binding protein (SLBP) on two threonines triggers degradation of SLBP, the sole cell cycle-regulated factor required for regulation of histone mRNA processing, at the end of S phase. *Mol Cell Biol*. 2003 Mar;23(5):1590-1601.
86. Sanchez R, Marzluff WF. The stem-loop binding protein is required for efficient translation of histone mRNA in vivo and in vitro. *Mol Cell Biol*. 2002 Oct;22(20):7093-7104.
87. Gorgoni B, Andrews S, Schaller A, et al. The stem-loop binding protein stimulates histone translation at an early step in the initiation pathway. *RNA*. 2005 Jul;11(7):1030-1042.
88. Huang Y, Gattoni R, Stevenin J, et al. SR splicing factors serve as adapter proteins for TAP-dependent mRNA export. *Mol Cell*. 2003 Mar;11(3):837-843.
89. Beggs S, James TC, Bond U. The PolyA tail length of yeast histone mRNAs varies during the cell cycle and is influenced by Sen1p and Rrp6p. *Nucleic Acids Res*. 2012 Mar;40(6):2700-2711.
90. Gunjan A, Verreault A. A Rad53 kinase-dependent surveillance mechanism that regulates histone protein levels in *S. cerevisiae*. *Cell*. 2003 Nov 26;115(5):537-549.
91. Navadgi-Patil VM, Burgers PM. Cell-cycle-specific activators of the Mec1/ATR checkpoint kinase. *Biochem Soc Trans*. 2011 Apr;39(2):600-605.
92. Singh RK, Kabbaj MH, Paik J, et al. Histone levels are regulated by phosphorylation and ubiquitylation-dependent proteolysis. *Nat Cell Biol*. 2009 Aug;11(8):925-933.
93. Rakesh Kumar Singh and Akash Gunjan. Histone tyrosine phosphorylation comes of age. *Epigenetics*. 2011 Feb; 6(2): 153-160.
94. Hanel W, Moll UM. Links between mutant p53 and genomic instability. *J Cell Biochem*. 2012 Feb;113(2):433-439.
95. Ubezio P, Lupi M, Branduardi D, et al. Quantitative assessment of the complex dynamics of G1, S, and G2-M checkpoint activities. *Cancer Res*. 2009 Jun 15;69(12):5234-5240.
96. Su C, Gao G, Schneider S, et al. DNA damage induces downregulation of histone gene expression through the G1 checkpoint pathway. *EMBO J*. 2004 Mar 10;23(5):1133-1143.
97. Mullen TE, Marzluff WF. Degradation of histone mRNA requires oligouridylation followed by decapping and simultaneous degradation of the mRNA both 5' to 3' and

- 3' to 5'. *Genes Dev.* 2008 Jan 1;22(1):50-65.
98. Emili A, Schieltz DM, Yates JR, 3rd, et al. Dynamic interaction of DNA damage checkpoint protein Rad53 with chromatin assembly factor Asf1. *Mol Cell.* 2001Jan;7(1):13-20.
99. Green EM, Antczak AJ, Bailey AO, et al. Replication-independent histone deposition by the HIR complex and Asf1. *Curr Biol.* 2005 Nov 22;15(22):2044-2049.
100. Sutton A, Bucaria J, Osley MA, et al. Yeast ASF1 protein is required for cell cycle regulation of histone gene transcription. *Genetics.* 2001 Jun;158(2):587-596.
101. Stephen P. Jackson and Jiri Bartek. The DNA-damage response in human biology and disease. *Nature.* 2009 Oct 22; 461(7267): 1071–1078.
102. Alexandre Maréchal and Lee Zou. DNA Damage Sensing by the ATM and ATR Kinases. *Cold Spring Harb Perspect Biol.* 2013 Sep; 5(9): a012716.
103. Helt CE, Cliby WA, Keng PC, et al. Ataxia telangiectasia mutated (ATM) and ATM and Rad3-related protein exhibit selective target specificities in response to different forms of DNA damage. *J Biol Chem.* 2005 Jan 14;280(2):1186-92.
104. Qianyun Mei, Junhua Huang, Shanshan Li, et al. Regulation of DNA replication-coupled histone gene expression. *Oncotarget.* 2017 Nov 7; 8(55): 95005–95022.
105. Tanae K, Horiuchi T, Matsuo Y, et al. Histone Chaperone Asf1 Plays an Essential Role in Maintaining Genomic Stability in Fission Yeast. *PLoS ONE* 7(1): e30472.
106. Jiao Y, Seeger K, Lautrette A, et al. Surprising complexity of the Asf1 histone chaperone-Rad53 kinase interaction. *Proc Natl Acad Sci U S A.* 2012 Feb 21;109(8):2866-71.
107. Berezney R, Dubey DD, Huberman JA. Heterogeneity of eukaryotic replicons, replicon clusters, and replication foci. *Chromosoma.* 2000 Mar;108(8):471-484.
108. Leonhardt H, Rahn HP, Weinzierl P, et al. Dynamics of DNA replication factories in living cells. *J Cell Biol.* 2000 Apr 17;149(2):271-280.
109. Paulovich AG, Hartwell LH. A checkpoint regulates the rate of progression through S phase in *S. cerevisiae* in response to DNA damage. *Cell.* 1995 Sep 8;82(5):841-847.
110. Santocanale C, Diffley JF. A Mec1- and Rad53-dependent checkpoint controls late-firing origins of DNA replication. *Nature.* 1998 Oct 8;395(6702):615-618.
111. Feijoo C, Hall-Jackson C, Wu R, et al. Activation of mammalian Chk1 during DNA replication arrest: a role for Chk1 in the intra-S phase checkpoint monitoring replication origin firing. *J Cell Biol.* 2001 Sep 3;154(5):913-923.
112. Tercero JA, Diffley JF. Regulation of DNA replication fork progression through damaged DNA by the Mec1/Rad53 checkpoint. *Nature.* 2001 Aug 2;412(6846):553-557.
113. Lengauer C, Kinzler KW, Vogelstein B. Genetic instabilities in human cancers. *Nature.* 1998 Dec 17;396(6712):643-649.
114. Ufuk Günesdogan, Herbert Jäckle, and Alf Herzig. Histone supply regulates S phase timing and cell cycle progression. *eLife.* 2014; 3: e02443.
115. Dunleavy EM, Almouzni G, Karpen GH. H3.3 is deposited at centromeres in S phase as a placeholder for newly assembled CENP-A in G₁ phase. *Nucleus.* 2011 Mar-Apr;2(2):146-57.
116. Reinberg D, Sims RJ, 3rd. de FACTo nucleosome dynamics. *J Biol Chem.* 2006 Aug 18;281(33):23297-23301.
117. Formosa T. FACT and the reorganized nucleosome. *Mol Biosyst.* 2008 Nov;4(11):1085-1093.
118. Orphanides G, LeRoy G, Chang CH, et al. FACT, a factor that facilitates transcript elongation through nucleosomes. *Cell.* 1998 Jan 9;92(1):105-116.
119. Pavri R, Zhu B, Li G, et al. Histone H2B monoubiquitination functions cooperatively with FACT to regulate elongation by RNA polymerase II. *Cell.* 2006 May 19;125(4):703-717.
120. Mason PB, Struhl K. The FACT complex travels with elongating RNA

- polymerase II and is important for the fidelity of transcriptional initiation in vivo. *Mol Cell Biol.* 2003 Nov;23(22):8323-8333.
121. Saunders A, Werner J, Andrulis ED, et al. Tracking FACT and the RNA polymerase II elongation complex through chromatin in vivo. *Science.* 2003 Aug 22;301(5636):1094-1096.
122. Jimeno-Gonzalez S, Gomez-Herreros F, Alepuz PM, et al. A gene-specific requirement for FACT during transcription is related to the chromatin organization of the transcribed region. *Mol Cell Biol.* 2006 Dec;26(23):8710-8721.
123. Stuwe T, Hothorn M, Lejeune E, et al. The FACT Spt16 "peptidase" domain is a histone H3-H4 binding module. *Proc Natl Acad Sci US A.* 2008 Jul 1;105(26):8884-8889.
124. VanDemark AP, Xin H, McCullough L, et al. Structural and functional analysis of the Spt16p N-terminal domain reveals overlapping roles of yFACT subunits. *J Biol Chem.* 2008 Feb 22;283(8):5058-5068.
125. Jamai A, Puglisi A, Strubin M. Histone chaperone spt16 promotes redeposition of the original h3-h4 histones evicted by elongating RNA polymerase. *Mol Cell.* 2009 Aug 14;35(3):377-383.
126. Morillo-Huesca M, Maya D, Munoz-Centeno MC, et al. FACT prevents the accumulation of free histones evicted from transcribed chromatin and a subsequent cell cycle delay in G1. *PLoS Genet.* 2010 May;6(5): e1000964.
127. Pan X, Ye P, Yuan DS, et al. A DNA integrity network in the yeast *Saccharomyces cerevisiae*. *Cell.* 2006 Mar 10;124(5):1069-1081.
128. Griffiths AJF, Miller JH, Suzuki DT, et al. An introduction to Genetic Analysis. 7th edition, WH Freeman publication. 2000.
129. Serra-Cardona A, Zhang Z. Replication-Coupled Nucleosome Assembly in the Passage of Epigenetic Information and Cell Identity. *Trends Biochem Sci.* 2018 Feb;43(2):136-148.
130. Varij nayan, Suneel kumar onteru, Dheer singh. Reproduction and nutriment-nurture crosstalk: An epigenetics perspective. *JRHM.* July 2015; 1:50-59.
131. R. Sanjuán, M. Pereira-Gómez, J. Risso. Genome Stability from virus to human application. Elsevier publication. 2016: p 37-47.
132. Douglas Maya, Macarena Morillo-Huesca, Lidia Delgado Ramos, et al. The Mechanisms of DNA Replication. IntechOpen publication. 2013: p 377-402.

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